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This document is the authors' final version of the published article.  
Link to published article: _http://dx.doi.org/10.1248/cpb.53.1480_

Recommended Citation  

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Identification and Comparative Determination of Senkyunolide A in Traditional Chinese Medicinal Plants Ligusticum chuanxiong and Angelica sinensis by HPLC Coupled with DAD and ESI-MS

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Received May 20, 2005; accepted August 2, 2005

Using the HPLC/DAD/ESI/MS method, the qualitative and quantitative analysis of senkyunolide A (SA) in the rhizomes of Ligusticum chuanxiong (Rhzoma chuanxiong; CX) and roots of Angelica sinensis (DG) was established. As a result, it was found that SA is a characteristic standard compound for the quality evaluation and chemical differentiation between CX and DG. Methanol was chosen in the preparation of standard solutions and extraction of samples based on the stability data. The identity of SA in CX and DG was unambiguously determined based on the quasimolecular ions in ESI-MS. A comprehensive validation of the method, including sensitivity, linearity, reproducibility and recovery, was conducted using the optimized chromatographic conditions. The linear calibration curve was acquired with $R^2$>0.999 and limit of detection (S/N=3) was estimated to be 12.5 $\mu$g/g. The reproducibility was evaluated by repeated sample injection and replicated analysis of samples with the relative standard deviation (RSD) value found within 0.68%. The recovery rates of SA varied within the range of 96.91—101.50% with RSD less than 2.38%. In the present work, the contents of SA were quantified within 3.94—9.14 mg/g and 0.108—0.588 mg/g for 12 batches each of CX and DG. The results demonstrated that SA is a useful standard compound for the quality evaluation and chemical differentiation between CX and DG. The analytical procedure is precise and reproducible and thus suitable for the analysis of a large number of samples.

Key words: senkyunolide A; Ligusticum chuanxiong; Angelica sinensis; HPLC/DAD/ESI/MS; Umbelliferae; quality evaluation

The present study mainly focuses on a precise, accurate quantitative method for comparison of SA in CX and DG with UV electrospray ionization (ESI) MS techniques.

Experimental

Materials and Reagents

Samples of CX and DG were collected in a number of cultivation bases in mainland China. The sources of the plant materials are listed in Table 1. The identities of these herbs were confirmed by appearance and microscopic and physiochemical analyses according to the Chinese Pharmacopoeia. Voucher specimens were deposited in the Herbarium Centre, Hong Kong Baptist University.

Reagent-grade solvents including petroleum ether (bp 35—60 °C), methanol, and ethyl acetate were purchased from Lab-Scan (Bangkok, Thailand) for the extraction of herbs and preparative TLC. HPLC-grade methanol (Lab-scan) was used as the mobile phase for HPLC. Deionized water was generated from a Milli-Q water system (Millipore, Bedford, MA, U.S.A.). Silica gel 60 F 254 preparative TLC plates (1.0 mm thickness, 20×20 cm, E. Merck, Darmstadt, Germany) were used for preliminary TLC identification. A semipreparative column (Supelcosil, PLC-18, 21.2 mm×250 mm, 12 $\mu$m, Supelco) was used in semipreparative HPLC isolation.

HPLC System and Conditions

An Agilent 1100 series HPLC-DAD system comprising a vacuum degasser, binary pump, autosampler, thermostated column compartment, and DAD (Hewlett Packard, U.S.A.) was used for quantitative analysis and UV spectra acquisition. The UV detector was set at the maximum absorption wavelength, i.e., 280 nm, of SA. For chromatographic analyses, an Alltima C18 column (5 $\mu$m, 4.6 mm×150 mm, Alltech Associates, Inc., U.S.A.) was used with a compatible guard column

![Fig. 1. Chemical Structures of (a) Ferulic Acid, (b) Z-ligustilide, and (c) Senkyunolide A (SA)](image)

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Notes

respectively. An aliquot of 10 ml of solution was injected for HPLC-DAD and HPLC-ESI-MS analyses. Sample duplicates were prepared as described above for analysis.

Results and Discussion

Optimization of the Isolation Process for SA Sonication was chosen as the extraction method in the present study. Observation of TLC showed that the amount of SA was more abundant while the polar impurities were minimized using petroleum ether as the extraction solvent. The extract was then subjected to preparative TLC to remove the major extraneous compounds and reduce the load capacity during subsequent semipreparative HPLC purification.

Optimization on the Preparation of SA Stock Solution and Sample Solution SA is a volatile oil under ambient conditions and therefore its stability remained a technical concern.6 For SA stock solution, its stability was evaluated by comparing the levels of SA in acetonitrile and methanol, respectively, over a period of storage. The levels of SA were found to be 98.7% and 98.3%, respectively, in acetonitrile and methanol after storing for 5 d, which indicated that SA was relatively stable in both solvent systems.

For the extraction of SA, sonication was chosen as the extraction method for its confirmed efficacy and ease of handling. The choice of extraction solvent for SA in herbs was further compared in methanol and acetonitrile. Persistent turbidity was observed in samples extracted with acetonitrile while the methanolic counterparts remained clear throughout. Petroleum ether was not considered owing to its incompatibility with the reverse-phase HPLC system. Therefore methanol was recommended as the solvent for preparation of standard solution and extraction of SA in herb samples.

Identification of SA in CX and DG by HPLC-MS
Apart from comparing with the retention time (tR), SA was further identified by HPLC-ESI-MS analysis to provide further information on its identity. Through comparing MS spectra acquired in negative- and positive-ion modes, the positive-ion mode was found to be more sensitive. For positive-ion mode MS spectra, consistent quasimolecular ions of SA, [M + H]+ (m/z = 193), [M + Na]+ (m/z = 215), and [M + K]+ (m/z = 231) were observed for peak S in HPLC chromatograms in standard solution, and CX and DG samples (Figs. 2a, b, c). These findings were in agreement with those reported in the literature.65

Calibration Curves and Limit of Detection SA was quantified in samples using the external standard addition method with a reference marker. Linearity was determined within the concentration range of 2.5—300 mg/l using nine standard solutions of different concentrations. Linear regression was expressed as Y = 4.885X + 2.37 with a correlation coefficient of 0.9998, where Y and X are the value of the area of peak and the concentration of standard solution, respectively. This regression equation was used for quantifying SA in all sample solutions.

The limit of detection (LOD) of SA in samples was determined based on visual evaluation with a signal-to-noise ratio of about 3 : 1. The LOD was estimated to be 0.25 mg/l in test solution, which was equivalent to 12.5 μg/g in solid samples.
Moreover, the quantitation limit of SA was determined based on a signal-to-noise ratio of about 10:1 for five replicated analyses of spiked matrix blank. The quantitation limit was found to be 0.95 mg/l in sample solutions, equivalent to 47.5 μg/g in solid samples. These results were considered satisfactory and acceptable for subsequent quantitative analysis.

**Method Reproducibility** Method reproducibility was evaluated with six repeated injections of standards and six replicated analysis of samples. The precision of replicated injections was determined and the relative standard deviation (RSD) of SA content was found to be 0.25% (n=6). The RSD of the content of SA in sample replicates was estimated to be 0.68% (n=6).

**Recovery** The recovery rate of SA was determined using spiked samples with different concentration levels of 50%, 100%, and 150% of SA in the samples. The recovery rates were estimated to be 101.50±1.54% (mean±RSD, n=3), 98.12±1.24% (mean±RSD, n=3), and 96.91±1.05% (mean±RSD, n=3), respectively. The mean recovery was 98.84±2.38% (n=9).

**Sample Analysis** The contents of SA in 12 batches of CX and 12 batches of DG are listed in Table 1. The contents of SA in CX and DG were within the ranges of 3.94—9.14 mg/g and 0.108—0.588 mg/g, respectively. On the other hand, the results showed that the content deviation of SA within the same medicinal plant material was also significant. This was probably attributed to the differences in cultivation conditions and processing methods, which gave rise to an inconsistent production of materia medica. However, the content of SA was generally more than 20-fold higher for CX than DG in the present study.

**Conclusion** In this study, the qualitative and quantitative analysis of SA in rhizomes of CX and roots of DG was established using
HPLC/DAD/ESI/MS. The overall procedure is rapid and reproducible and thus suitable for the analysis of numerous samples.

From the results, the content of SA in CX was generally 20-fold greater than in DG. Therefore SA is useful as a characteristic standard compound for the quality evaluation as well as chemical differentiation between these two closely related umbelliferic medicinal plants.

Acknowledgment The authors would like to thank Mr. Zhonghui Li (Department of Chemistry, Hong Kong Baptist University) for technical assistance in the NMR analysis.

References